



Assembly of Bacterial Genome Sequences from Metagenomes of Spacecraft Assembly Cleanrooms

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ABSTRACT Characterizing the microbiome of spacecraft assembly cleanrooms is important for planetary protection. We report two bacterial metagenome-assembled genomes (MAGs) reconstructed from metagenomes produced from cleanroom samples from the Kennedy Space Center's Payload Hazardous Servicing Facility (KSC-PHSF) during the handling of the Phoenix spacecraft. Characterization of these MAGs will enable identification of the strategies underpinning their survival.

To avoid microbial contamination during planetary missions, there are standards for spacecraft bioburden that are maintained by the Committee on Space Research (COSPAR) Planetary Protection Panel (<https://cosparhq.cnes.fr/scientific-structure/panels/panel-on-planetary-protection-ppp/>). Regulations require that spacecraft be assembled in cleanrooms with defined sterilization protocols (1). Studies characterizing these environments have identified a persistent microbiome (1–5). One such study characterized the microbiome of the Kennedy Space Center's Payload Hazardous Servicing Facility (KSC-PHSF) during the assembly of the Phoenix spacecraft (1).

In that study by Bashir et al. (1), DNA samples were collected with a biological sampling kit (BiSKits, QuickSilver Analytics) from 1 m² of the floor of the KSC-PHSF during (8 samples) and after (10 samples) the spacecraft assembly (1). DNA extraction was performed using bead beating and an automated DNA extraction instrument (Autolyser A-2 DNA automated platform, Axcyte Genomics). Samples from each sampling period were pooled following extraction. Each sample was amplified using multiple displacement amplification with a REPLI-g single-cell whole-genome amplification kit (Qiagen). DNA was sheared using an E210 instrument (Covaris, Woburn, MA) and then end repaired, A tailed, and ligated to Illumina adaptors according to standard Illumina (San Diego, CA) paired-end (PE) protocols. Metagenomic sequencing of the materials was performed on a HiSeq 2500 sequencing instrument (Illumina) with paired-end 2 × 250-bp read lengths (1). This produced 4,654,014 and 22,355,430 raw reads for the metagenomes collected during spacecraft assembly and after spacecraft assembly, respectively. In this study, default parameters were used for all software. The raw reads were downloaded from NCBI GenBank, and the short and poor-quality sequences were excluded using Trimmomatic v0.39 (6). The reads were assembled into contigs using MEGAHIT v1.1.3 (7) (after spacecraft assembly: 114,901 contigs; N_{50} , 401 bp; during spacecraft assembly: 12,199 contigs; N_{50} , 1,130 bp). Contigs were binned into metagenome-assembled genomes (MAGs) using MaxBin v2.2.7 (8). The taxonomic classification, completeness, and contamination of these MAGs were assessed using CheckM v1.1.2 (9). A medium-quality MAG was produced from the metagenome collected during spacecraft assembly (MAG-P1) and a high-quality MAG (MAG-P2) from the metagenome collected after spacecraft assembly (10). The

Citation Ilieva V, Steel B, Pratscher J, Olsson-Francis K, Macey MC. 2021. Assembly of bacterial genome sequences from metagenomes of spacecraft assembly cleanrooms. *Microbiol Resour Announc* 10:e01439-20. <https://doi.org/10.1128/MRA.01439-20>.

Editor J. Cameron Thrash, University of Southern California

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Received 18 January 2021

Accepted 29 January 2021

Published 18 February 2021

contamination scores for MAG-P1 were improved using VizBin v1.0.0 (11). tRNAs and rRNA genes were identified using Aragorn v1.2.38 (12) and RNAmmer v1.2 (13).

The high-quality MAG was classified as *Rhizobium* (MAG-P2), with completeness and contamination scores of 96.33% and 4.69%, respectively, and 438-fold coverage. MAG-P2 is composed of 468 contigs and contains 1 16S rRNA gene copy, 64 tRNAs, and 7,771 coding sequences (CDSs). The genome size is 8.36 Mb, with 63.1% GC content. CheckM classified MAG-P1 in the genus *Acinetobacter*, with completeness and contamination scores of 52.90% and 8.62%, respectively, and 863-fold coverage. MAG-P1 is 4.34 Mb, with 39.8% GC content, 1,793 contigs, and 4,253 CDSs.

Members of *Rhizobiaceae* and *Acinetobacter* were previously detected in the KSC-PHSF, as well as other cleanroom environments (1, 5, 14–20). Further analysis of these MAGs will provide insight into the strategies underpinning survival in cleanroom environments and inform future sterilization strategies.

Data availability. The sequences analyzed in this study are available from the NCBI Sequence Read Archive under accession numbers [SRX1896153](https://www.ncbi.nlm.nih.gov/sra/SRX1896153) and [SRX1896154](https://www.ncbi.nlm.nih.gov/sra/SRX1896154). The whole-genome shotgun projects were deposited at DDBJ/ENA/GenBank under accession numbers [JAEPRK000000000](https://www.ncbi.nlm.nih.gov/nuccore/JAEPRK000000000) (*Acinetobacter* sp. strain MAG-P1) and [JAEPLR000000000](https://www.ncbi.nlm.nih.gov/nuccore/JAEPLR000000000) (*Rhizobium* sp. strain MAG-P2). The versions described in this paper are the first versions.

ACKNOWLEDGMENTS

The analysis described in the manuscript was supported by the Research England Expanding Excellence in England (E3) fund (124.18), an STFC consolidated grant (ST/T000228/1), a Natural Environment Research Council (NERC) independent research fellowship (NE/L010771/2), and a NERC EnvEast grant. The original research was funded by Planetary Protection Research program in NNH11ZDA001N, ROSES 2011 awarded to Parag A. Vaishampayan, and part of the research was carried out at the Jet Propulsion Laboratory, California Institute of Technology, under contract with the National Aeronautics and Space Administration.

We thank Parag A. Vaishampayan for his support during our writing of the manuscript and the research team of the original paper.

REFERENCES

- Bashir M, Ahmed M, Weinmaier T, Ciobanu D, Ivanova N, Pieber TR, Vaishampayan PA. 2016. Functional metagenomics of spacecraft assembly cleanrooms: presence of virulence factors associated with human pathogens. *Front Microbiol* 7:1321. <https://doi.org/10.3389/fmicb.2016.01321>.
- Zhang Y, Zhang L-T, Li Z-D, Xin C-X, Li X-Q, Wang X, Deng Y-L. 2019. Microbiomes of China's space station during assembly, integration, and test operations. *Microb Ecol* 78:631–650. <https://doi.org/10.1007/s00248-019-01344-4>.
- Weinmaier T, Probst AJ, La Duc MT, Ciobanu D, Cheng J-F, Ivanova N, Rattei T, Vaishampayan P. 2015. A viability-linked metagenomic analysis of cleanroom environments: eukarya, prokaryotes, and viruses. *Microbiome* 3:62. <https://doi.org/10.1186/s40168-015-0129-y>.
- Koskinen K, Rettberg P, Pukall R, Auerbach A, Wink L, Barczyk S, Perras A, Mahnert A, Margheritis D, Kminek G, Moissl-Eichinger C. 2017. Microbial biodiversity assessment of the European Space Agency's ExoMars 2016 mission. *Microbiome* 5:143. <https://doi.org/10.1186/s40168-017-0358-3>.
- Mahnert A, Vaishampayan P, Probst AJ, Auerbach A, Moissl-Eichinger C, Venkateswaran K, Berg G. 2015. Cleanroom maintenance significantly reduces abundance but not diversity of indoor microbiomes. *PLoS One* 10:e0134848. <https://doi.org/10.1371/journal.pone.0134848>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Li D, Liu C-M, Luo R, Sadakane K, Lam T-W. 2015. MEGAHIT: an ultra-fast single-node solution for large and complex metagenomics assembly via succinct de Bruijn graph. *Bioinformatics* 31:1674–1676. <https://doi.org/10.1093/bioinformatics/btv033>.
- Wu Y-W, Simmons BA, Singer SW. 2016. MaxBin 2.0: an automated binning algorithm to recover genomes from multiple metagenomic datasets. *Bioinformatics* 32:605–607. <https://doi.org/10.1093/bioinformatics/btv638>.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
- Bowers RM, Kyrpides NC, Stepanauskas R, Harmon-Smith M, Doud D, Reddy TBK, Schulz F, Jarett J, Rivers AR, Eloe-Fadrosh EA, Tringe SG, Ivanova NN, Copeland A, Clum A, Becraft ED, Malmstrom RR, Birren B, Podar M, Bork P, Weinstock GM, Garrity GM, Dodsworth JA, Yooseph S, Sutton G, Glöckner FO, Gilbert JA, Nelson WC, Hallam SJ, Jungbluth SP, Etema TJG, Tighe S, Konstantinidis KT, Liu W-T, Baker BJ, Rattei T, Eisen JA, Hedlund B, McMahon KD, Fierer N, Knight R, Finn R, Cochrane G, Karsch-Mizrachi I, Tyson GW, Rinke C, Lapidus A, Meyer F, Yilmaz P, Parks DH, Eren AM, The Genome Standards Consortium, et al. 2017. Minimum information about a single amplified genome (MISAG) and a metagenome-assembled genome (MIMAG) of bacteria and archaea. *Nat Biotechnol* 35:725–731. <https://doi.org/10.1038/nbt.3893>.
- Laczny CC, Sternal T, Plugaru V, Gawron P, Atashpendar A, Margossian HH, Coronado S, van der Maaten L, Vlassis N, Wilmes P. 2015. VizBin—an application for reference-independent visualization and human-augmented binning of metagenomic data. *Microbiome* 3:1–7. <https://doi.org/10.1186/s40168-014-0066-1>.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <https://doi.org/10.1093/nar/gkh152>.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt H-H, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 35:3100–3108. <https://doi.org/10.1093/nar/gkm160>.
- Vaishampayan P, Osman S, Andersen G, Venkateswaran K. 2010. High-density 16S microarray and clone library-based microbial community composition of the Phoenix spacecraft assembly clean room. *Astrobiology* 10:499–508. <https://doi.org/10.1089/ast.2009.0443>.
- La Duc MT, Vaishampayan P, Nilsson HR, Torok T, Venkateswaran K. 2012.

- Pyrosequencing-derived bacterial, archaeal, and fungal diversity of spacecraft hardware destined for Mars. *Appl Environ Microbiol* 78:5912–5922. <https://doi.org/10.1128/AEM.01435-12>.
16. Ghosh S, Osman S, Vaishampayan P, Venkateswaran K. 2010. Recurrent isolation of extremotolerant bacteria from the clean room where Phoenix spacecraft components were assembled. *Astrobiology* 10:325–335. <https://doi.org/10.1089/ast.2009.0396>.
 17. Checinska A, Probst AJ, Vaishampayan P, White JR, Kumar D, Stepanov VG, Fox GE, Nilsson HR, Pierson DL, Perry J, Venkateswaran K. 2015. Microbiomes of the dust particles collected from the International Space Station and spacecraft assembly facilities. *Microbiome* 3:50. <https://doi.org/10.1186/s40168-015-0116-3>.
 18. Seuylemezian A, Cooper K, Schubert W, Vaishampayan P. 2018. Draft genome sequences of 12 dry-heat-resistant *Bacillus* strains isolated from the cleanrooms where the Viking spacecraft were assembled. *Genome Announc* 6:e00094-18. <https://doi.org/10.1128/genomeA.00094-18>.
 19. Vaishampayan P, Probst AJ, La Duc MT, Bargoma E, Benardini JN, Andersen GL, Venkateswaran K. 2013. New perspectives on viable microbial communities in low-biomass cleanroom environments. *ISME J* 7:312–324. <https://doi.org/10.1038/ismej.2012.114>.
 20. Mogul R, Barding GA, Jr, Lalla S, Lee S, Madrid S, Baki R, Ahmed M, Brasali H, Cepeda I, Gornick T, Gunadi S, Hearn N, Jain C, Kim EJ, Nguyen T, Nguyen VB, Oei A, Perkins N, Rodriguez J, Rodriguez V, Savla G, Schmitz M, Tedjakesuma N, Walker J. 2018. Metabolism and biodegradation of spacecraft cleaning reagents by strains of spacecraft-associated *Acinetobacter*. *Astrobiology* 18:1517–1527. <https://doi.org/10.1089/ast.2017.1814>.