Ultrastructural Distribution of the 7 Nicotinic Acetylcholine Receptor Subunit in Rat Hippocampus

How to cite:

For guidance on citations see FAQs.

© [not recorded]

Version: [not recorded]

Link(s) to article on publisher’s website:
http://www.jneurosci.org/cgi/content/abstract/21/20/7993?maxtoshow=&HITS=10&hits=10&RESULTFORMAT=&author1=skehel&andorexactfulltext=and&searchid=1&FIRSTINDEX=0&sortspec=relevance&resourcetype=HWCIT

Copyright and Moral Rights for the articles on this site are retained by the individual authors and/or other copyright owners. For more information on Open Research Online’s data policy on reuse of materials please consult the policies page.

oro.open.ac.uk
Acetylcholine (ACh) is an important neurotransmitter in the mammalian brain; it is implicated in arousal, learning, and other cognitive functions. Recent studies indicate that nicotinic receptors contribute to these cholinergic effects, in addition to the established role of muscarinic receptors. In the hippocampus, where cholinergic involvement in learning and memory is particularly well documented, α7 nicotinic acetylcholine receptor subunits (α7 nAChRs) are highly expressed, but their precise ultrastructural localization has not been determined. Here, we describe the results of immunogold labeling of serial ultrathin sections through stratum radiatum of area CA1 in the rat. Using both anti-α7 nAChR immunolabeling and α-bungarotoxin binding, we find that α7 nAChRs are present at nearly all synapses in CA1 stratum radiatum, with immunolabeling present at both presynaptic and postsynaptic elements. Morphological considerations and double immunolabeling indicate that GABAergic as well as glutamatergic synapses bear α7 nAChRs, at densities approaching those observed for glutamate receptors in CA1 stratum radiatum. Postsynaptically, α7 nAChRs often are distributed at dendritic spines in a perisynaptic annulus. In the postsynaptic cytoplasm, immunolabeling is associated with spine apparatus and other membranous structures, suggesting that α7 nAChRs may undergo dynamic regulation, with insertion into the synapse and subsequent internalization. The widespread and substantial expression of α7 nAChRs at synapses in the hippocampus is consistent with an important role in mediating and/or modulating synaptic transmission, plasticity, and neurodegeneration.

Key words: acetylcholine receptors; nicotine; dendritic spines; postsynaptic density; immuno-electron microscopy; glutamate; GABA; Aβ1–42

Received Feb. 21, 2001; revised June 13, 2001; accepted June 28, 2001.

This work was supported by the Medical Research Council and by grants from the Human Frontier Science Program (A.F.) and the Brain and Behavioural Sciences Research Council (108/BI 11211, M.G.S.). We thank J.-A. Horne, E. Hirst, and Drs. T. V. F. Bliss, I. Burdett, and I. A. Meinertzhagen for helpful discussion. Correspondence should be addressed to Dr. Ruth Fabian-Fine, Department of Psychology, Dalhousie University, Halifax, Nova Scotia B3H 4J1 Canada. E-mail: rfabian@is.dal.ca.

Copyright © 2001 Society for Neuroscience 0270-6474/01/217993-11$15.00/0


Ultrastructural Distribution of the α7 Nicotinic Acetylcholine Receptor Subunit in Rat Hippocampus

Ruth Fabian-Fine,1,2 Paul Skehel,2 Mick L. Errington,2 Heather A. Davies,1 Emanuele Sher,3 Michael G. Stewart,1, and Alan Fine2,4

1Department of Biological Sciences, The Open University, Milton Keynes, MK7 6AA, United Kingdom, 2Division of Neurophysiology, National Institute for Medical Research, Mill Hill, London NW7 1AA, United Kingdom, 3Lilly Research Centre, Eli Lilly and Co., Erl Wood Manor, Windlesham, Surrey GU20 6PH, United Kingdom, and 4Department of Physiology and Biophysics, Dalhousie University Faculty of Medicine, Halifax, Nova Scotia B3H 4H7, Canada
post synaptic nature of the labeling. Electron microscopic (EM) analysis of 125I-α-Bgtx binding provided evidence for α7 nAChRs at hippocampal synapses (Hunt and Schmidt, 1978), but the large grain radius and relative insensitivity of the method prevented firm conclusions about incidence or distribution of the α-Bgtx binding sites. Full understanding of the varied and subtle functional roles recently attributed to α7 nAChRs in the hippocampus (Radcliffe et al., 1999) will require high-resolution analysis of the subcellular distribution of this subunit. To this end, we have performed light and electron microscopic immunostaining of CA1 stratum radiatum, and here report that α7 receptors are highly abundant at almost all synapses in this region. The intensity of the signal suggests that the importance of α7-mediated nicotinic cholinergic signaling may be far greater than is currently recognized.

MATERIALS AND METHODS

Light microscopic α7 nAChR immunolabeling. Intact brain preparations were obtained from two adult male Sprague Dawley rats. Animals were anesthetized with urethane (1.5 gm/kg, i.p.) and perfused over ~20 min with 200 ml of 4% paraformaldehyde (PFA)/0.3% glutaraldehyde (GA) in PBS (0.1 M, pH 7.2). After dissection the brains were immersed in the same fixative for 2 hr and embedded in 7% Agarose. Vibratome sections 30 μm thick were cut transversely through the brain using a Leica VT1000S and rinsed in PBS (4 × 10 min). Sections containing the hippocampal formation were then incubated in 1% glycine in PBS to quench residual reactive aldehyde groups, washed in PBS (2 × 10 min), permeabilized in 0.1% saponin (S2149; Sigma, St. Louis, MO)/PBS for 15 min, and incubated with the primary monoclonal anti-α7 nAChR antibody Mab 306 (M220, Sigma) (Schoepfer et al., 1990) at 4°C overnight. The antibody was diluted 1:3000 in an incubation medium (IM) consisting of PBS with 1% bovine serum albumin (A5403, Sigma), 5% normal goat serum, 0.5% cold water fish skin gelatin (G7765, Sigma), and 0.01% saponin. After incubation, preparations were rinsed thoroughly in PBS. To detect anti-α7 nAChR antibody binding, a secondary goat anti-mouse antibody coupled to the fluorochrome Cy3 (Stratec Scientif, Luton, UK; 115-165-003; 1:500 in IM at 4°C overnight) was used. Preparations were then washed in PBS (6 × 1 hr), mounted in Mowiol 4–88 (475904, Calbiochem, La Jolla, CA) on glass slides, and examined using a Zeiss Axiopt microscope with epifluorescence optics. Red Cy3-immunofluorescence was detected using a standard rhodamine filter set. Preparations were immediately photographed on color slide film (Ektachrome 400). Light microscopic labeling for synaptophysin and α-Bgtx. Hippocampal preparations were obtained from two adult male Sprague Dawley rats. Animals, anesthetized as above, were perfused over ~10 min with 200 ml of freshly prepared 4% PFA in PBS. Brains were removed, embedded in 7% Agarose, and 20 μm vibratome sections obtained as above. After permeabilization with 0.01% saponin in IM (30 min), sections were incubated in IM containing a monoclonal anti-synaptophysin antibody (1:120; SY38, 902322; Boehringer Mannheim, Mannheim, Germany) overnight at 4°C. After washing in PBS (5 × 1 hr) and 0.01% saponin in IM, preparations were incubated overnight at 4°C in IM containing a secondary goat anti-mouse antibody coupled to Cy3 (1:500; see above) and Alexa488-conjugated α-Bgtx (1:500; B13422; Molecular Probes). After thorough rinsing in PBS (4 × 1 hr), slices were mounted in Mowiol and examined with a Leica TCS NT confocal microscope using standard fluorescein and rhodamine filter combinations.

Postembedding immunogold labeling for electron microscopy. Freeze-substituted tissue was used to preserve greater immunoreactivity of the tissue and achieve maximum staining intensity. Three adult (all 22 months old) CFHB male rats were anesthetized with Sagatal and perfused through the heart with 50 ml of 0.9% saline followed by 250 ml 0.1 M phosphate buffer, pH 7.4, containing 4% paraformaldehyde, 0.05% glutaraldehyde, and 0.2% picric acid (Somogyi and Takagi, 1982). Hippocampus was dissected immediately, and ~0.5 mm slabs were cut by hand from the dorsal part, soaked in PB for 15 min, and then immersed in 0.125% triethanolamine hydrochloride in PB for 30 min to quench unreacted aldehydes. The tissue slabs were cryoprotected over 2 hr in increasing concentrations of glycerol in PBS to 30% and then impact-frozen on a polished copper mirror at ~193°C in a Leica MM80 impact freezer. Freeze-sliced slabs were treated ex situ in a Leica AFS automatic freeze substitution system in methanol with 0.5% uranyl acetate for 24 hr at ~85°C, rinsed in methanol, and the temperature raised to ~50°C before infiltration and embedding in Lowicryl HM 20 (R1034; Agar Scientific, Stansted, England). The resin was polymerized at ~50°C by exposure to ultraviolet light. Serial ultrathin sections (50 nm) were cut with a Reichert Ultracut and collected on pioloform-coated single-slot nickel grids. To treat all sections identically and to prevent tearing during the labeling procedure, grids were mounted in a grid support plate (16705698; Leica Microsystems, Milton Keynes, UK). Sections were wetted in PBS for 30 min and preincubated in IM for 30 min at room temperature. Sections were then incubated with the monoclonal anti-α7 nAChR antibody (see above; 1:4000 in IM) overnight at 4°C followed by 1 hr at 37°C. After thorough washing in PBS and preincubation in IM (30 min), secondary antibodies were applied at a dilution of 1:100 in IM containing mouse anti-biotin (1:100 in IM; SC5544; Santa Cruz Biotechnology, CA) detected with goat anti-mouse 10 nm gold biotin-XX-conjugated (A-11242; Molecular Probes). AlexaFluor568-conjugated α-Bgtx (1:500; B-1196; Molecular Probes). After thorough rinsing in PBS, sections were incubated overnight at 4°C in IM containing mouse anti-biotin antibody (1:250; A-11242; Molecular Probes). For detection of the mouse anti-biotin antibody, a goat anti-mouse antibody coupled to 10 nm gold was used

Figure 1. Distribution of α7 nAChR-LIR in rat hippocampus. A, Low magnification micrograph of the hippocampal formation shows that all areas display α7 nAChR-LIR. The strongest immunoreactivity is present in the cell body layers (1, dentate gyrus; 2, CA3; 3, CA1) and in the molecular layer of the dentate gyrus (arrows). B, C, Higher magnification of the CA3 (B) and CA1 (C) regions show that the apical dendrites of the pyramidal cells (arrowheads) display α7 nAChR-LIR. Scale bar: A, 1 mm; B, C, 170 μm.
Mito

nAChR α7 i

nAChR α7 ii

Hsp-60

Figure 2. A, The specificity of the mitochondrial labeling seen with immuno-electron microscopy was addressed by Western blot analysis of hippocampal proteins. The monoclonal anti-α7 nAChR antibody M220 labels two bands in the homogenate and low-speed pellet, with molecular masses of 56 and 46 kDa (nAChRa7, i). The 56 kDa protein, which corresponds to the α7 nAChR subunit, is not present in the mitochondrial fraction (Mito). The 46 kDa immunoreactivity is sensitive to binding conditions and is not present when the binding buffer contains 3% nonfat milk powder (nAChRa7, ii). The mitochondrial protein Hsp-60 is enriched in the mitochondrial fraction. B, Electron micrographs of the mitochondrial fraction tested in A demonstrate that these organelles were highly enriched in the pellet (arrowheads). C–E, Double labeling of different epitopes of α7 nAChR with the monoclonal antibody M220 (5 nm particles) and a polyclonal antiserum SC5544 (10 nm particles) demonstrates that both reagents label synaptic sites (+) and are often colocalized (arrows). Scale bar: B, 370 nm; C–E, 280 nm.

(1:100; 4 hr at 37°C; see above). The labeling in control preparations in which the biotin-XX-conjugated aBt was omitted did not exceed background intensity and revealed no evidence that the biotin binds unspecifically to synaptic sites.

Mitochondria preparation and Western blot analysis. Mitochondria were prepared from the hippocampus of four 115 gm male Sprague Dawley rats, according to standard methods (Lavrump and Zelander, 1982). Briefly, hippocampus was homogenized on ice in 10 vol of 0.44 M sucrose, 10 mM HEPES, and 1 mM MgCl₂. The homogenate was then centrifuged at 2000 rpm (400 × g) for 10 min at 4°C in a Beckman JA-20. The supernatant was removed, leaving P1, and centrifuged again at 14,000 rpm (17,500 × g) for 15 min under the same conditions. The resulting supernatant S2 was removed, and the pellet was resuspended in ice-cold homogenization buffer and centrifuged again at 9000 rpm (7000 × g) for 15 min to generate a supernatant S3 and a pellet enriched for mitochondria. This operation was repeated twice more to wash the mitochondrial fraction. Equivalent amounts of each fraction and the initial homogenate were analyzed by Western blot analysis. Proteins were separated by SDS-PAGE as described previously (Schägger and von Jagow, 1987) using a 10% separating gel, and transferred to Immobilon-P by electroblotting. Membranes were blocked in either 2% (w/v) BSA (Fraction V, 735086, Boehringer Mannheim), 0.05% (v/v) Nonidet P40 (BDH, Poole UK) in PBS, or 3% nonfat milk powder, 0.05% (v/v) NP40 in PBS. Primary antibodies were applied in the same buffer at dilutions of 1:10,000 for anti-α7 nAChR (M220) and 1:30,000 for anti-Hsp60 (SPA-804; StressGen, Victoria British Columbia, Canada). Immunoreactivity was detected using HRP-conjugated donkey anti-rabbit (711-035-152; Stratech Scientific, Luton, UK) or anti-mouse (715-035-150; Stratech Scientific) serum at 1:10,000 and ECL (RPN 2106, Amersham Pharmacia Biotech, Little Chalfont, UK).

Controls. The specificity of the mouse anti-α7 nAChR antibodies used here has been described previously (Schoepfer et al., 1990; Dominguez del Toro et al., 1994). Specificity of antibody binding was confirmed by immunoblots and by the absence of immunolabeling in preparations from which primary antibodies were omitted. The anti-synaptophysin antibody used here is well characterized and is known to bind selectively to presynaptic terminals close to the synaptic cleft; and (3) presence of a postsynaptic density in the postsynaptic profile. According to the appear-
ance of the postsynaptic density, two different synapse types have been
distinguished in the vertebrate CNS (Gray, 1959; Colonnier, 1968). The
first type (type 1 or "asymmetrical synapse") has an extensive postsyn-
aptic density and a population of large, round, electron-lucent vesicles in
the presynaptic profile. Type 2 ("symmetrical synapses") are character-
ized by a less conspicuous postsynaptic density and a presynaptic popu-
lation of small, pleomorphic, electron-lucent vesicles. It is generally
accepted that glutamatergic synapses have asymmetric morphology,
whereas GABAergic synapses are symmetric. Complete counts of all
gold particles at each synapse would require full three-dimensional
reconstruction of each synapse, a prohibitively time-consuming task. As
a useful approximation that largely eliminates false-negative (unlabeled)
synapses, we analyzed sets of three serial sections. All synapses in
randomly selected fields of view that were present throughout all three
serial sections were counted. For presentation, micrographs and slides
were digitized at high resolution using a Mustek flatbed scanner or a
Nikon LS-1000 slide scanner. Contrast and brightness were optimized in
Adobe Photoshop 5.0.

Statistical analysis. Gold particle incidence was compared to determine
whether the α7 nAChR immunolabeling over synaptic membranes was
significantly higher than background labeling. Gold particles were
counted over the synaptic membranes identified in all serial sections. The
actual sampled region was the synaptic cleft plus the adjoining 30-nm-
wide presynaptic and postsynaptic zones, because the separation between
a labeled epitope and a gold particle attached to a secondary antibody can
extend up to 28 nm (Matsubara et al., 1996). These counts were com-
pared with the number of gold particles over equivalent areas of regions
where no labeling was expected, such as myelin. Because particle counts
were not normally distributed, a Welch t test was used for comparisons.
The sampled synaptic area for each synapse was estimated by multiplying
the total sampled length of the synaptic cleft over the three serial sections
by the section thickness. Linear regression analysis was performed to
examine possible correlation of the number of gold particles per synapse
with the synaptic area.

Figure 3. A–C, Light microscopic double labeling for αBgtx (green) and synaptophysin (red) demonstrates the abundance and substantial correspon-
dence of both labeled sites. D, Higher magnification of the area outlined in C reveals the predominant colocalization (arrow) or close apposition of both
labeled sites. A small proportion of synaptophysin-labeled sites (double arrowhead) and a fraction of αBgtx-labeled sites (arrowhead) show no
colocalization. Scale bar: A–C, 130 μm; D, 30 μm.

Figure 4. Electron micrographs of the CA1 stratum radiatum area in
αBgtx-labeled intact brain preparations. A–E, Labeling for αBgtx is
present at presynaptic (arrowheads) and postsynaptic sites (arrows). b,
Presynaptic boutons; s, dendritic spines. Scale bar, 200 nm.
RESULTS
Light and electron microscopic immunolabeling demonstrates that the α7 nAChR subunit is present in cell bodies and processes of hippocampal neurons

Light microscopic immunolabeling of the hippocampal formation revealed diffuse α7 nAChR-like immunoreactivity (α7 nAChR-LIR) throughout cell bodies and cell processes of neurons in the dentate gyrus and CA3 and CA1 regions (Fig. 1). Immunoreactivity throughout the molecular layer of the dentate gyrus and in the cell body layers of all regions was relatively strong and easily recognizable at low magnification (Fig. 1A). Immunoreactivity in the dendritic fields of CA3 and CA1 was weaker, but clearly visible at higher magnification in Figure 1, B and C. To determine the precise location of the α7 nAChRs, we performed postembedding EM immunolabeling. Synaptic contacts were heavily labeled, as were membranous structures within the presynaptic and postsynaptic cytoplasm. Labeling was also present at mitochondria (Fig. 2); to determine whether this labeling represented authentic α7 nAChRs or cross-reactivity of the antibody with other protein(s) present in mitochondria, we performed Western blot analysis on subcellular fractions of brain homogenates. Our results demonstrate that the monoclonal antibody M220 recognizes two bands in crude homogenates, one of ~56 kDa (corresponding to the α7 nAChR) and one of 44 kDa (Fig. 2A). Only the 44 kDa band was present in the purified mitochondrial fraction. That this fraction contained mainly mitochondria was con-

Figure 5. Electron micrographs of the CA1 stratum radiatum region in anti-α7 nAChR-labeled intact brain preparations. A–F. Most synaptic contacts contain numerous gold particles at synaptic membranes (arrowheads). Gold particles are also found at presynaptic vesicles (small arrow) and attached to membranous structures in the postsynaptic cytoplasm (double arrowheads). Inset, Gold particles were often found at nonsynaptic membranes at positions corresponding to synapses in adjacent sections (arrowheads), indicating a perisynaptic localization of the α7 nAChR subunits. b, Presynaptic boutons; s, dendritic spines. Scale bar: A–F, 200 nm; inset, 90 nm.
confirmed by its enrichment in the mitochondrial marker, Hsp-60 (Fig. 2A), and by ultrastructural investigation of the fraction (Fig. 2B). We were able to eliminate the 44 kDa band in Western blots using 3% nonfat milk powder in the blocking medium (Fig. 2A) (see Materials and Methods). This blocking procedure, however, was not compatible with immuno-electron microscopy. To confirm the specificity of α7 nAChR immunolabeling, we performed two independent tests: (1) double labeling using the monoclonal antibody (M220) and a polyclonal antiserum (SC5544) raised against a larger epitope of the α7 subunit (polyclonal: amino acids 367–502; monoclonal: amino acids 380–400); and (2) light and electron microscopic labeling for αBgtx. Figure 2C–E demonstrate that both antibodies directed against the α7 subunit labeled synaptic contacts, frequently colocalizing at both presynaptic and postsynaptic sites. As shown in Figure 2, D and E (5 nm particles), the monoclonal antibody yielded stronger labeling, and only the monoclonal antibody labeled mitochondria (Fig. 2C–E).

The presence of α7 nAChR at synaptic sites was further confirmed by αBgtx binding. As demonstrated in Figure 3, light microscopic double labeling for αBgtx and synaptophysin revealed abundant expression of both epitopes in hippocampal synaptic zones, often in close apposition. In keeping with previous findings (Hunt and Schmidt, 1979), αBgtx binding was seen only occasionally throughout neuronal cell bodies; this difference compared with the pattern of anti-α7 nAChR immunolabeling may reflect the post-translational processing required for αBgtx binding (Chen et al., 1998; Aztiria et al., 2000) but not for binding of the monoclonal antibody directed against the peptide epitope. Ultrastructural investigation of hippocampal tissue labeled for αBgtx revealed widespread presynaptic and postsynaptic labeling (Fig. 4) that, although less intense than the labeling observed with the anti-α7 nAChR antibody, was qualitatively similar in distribution, confirming that the synaptic immunolabeling reflects the presence of authentic α7 nAChRs.

### Most synapses in CA1 stratum radiatum display α7 nAChR-LIR

The ultrastructural investigation of dentate gyrus and CA1 and CA3 regions revealed α7 nAChR-LIR at synaptic sites in all hippocampal areas (Fig. 5). Gold particles were located mainly (1) at the postsynaptic density, (2) within the synaptic cleft, (3) in the postsynaptic cytoplasm (Figs. 5–7), and (4) at presynaptically located vesicles, often located in close proximity to the synaptic contacts.
membrane but >30 nm apart from it (indicating that the epitope is located at the vesicles rather than the synaptic membrane). It is not clear whether the latter are synaptic vesicles or specialized transport vesicles. Labeled synapses were present at both den-
glutamatergic and virtually all synapses showed \( \alpha 7 \) nAChR-LIR, most glutamatergic synapses are likely to bear substantial levels of \( \alpha 7 \) nAChRs. Consistent with this interpretation, double labeling for glutamate and \( \alpha 7 \) nAChR shows that most but not all asymmetric \( \alpha 7 \) nAChR-LIR synapses are glutamate-LIR (Fig. 9); glutamate-LIR, \( \alpha 7 \) nAChR-negative synapses were rare. Double labeling for both \( \alpha 7 \) nAChRs/GABA and \( \alpha 7 \) nAChR/glutamate also revealed that most of the postsynaptic neurons with \( \alpha 7 \) nAChRs were glutamatergic, presumably pyramidal cells.

To investigate whether the \( \alpha 7 \) nAChR-immunoreactive contacts include GABAergic synapses, we performed double labeling for GABA and \( \alpha 7 \) nAChRs. The results show that many but not all GABA-like immunoreactive profiles display \( \alpha 7 \) nAChR-LIR at the synaptic cleft (Fig. 10), suggesting that many GABAergic synapses in CA1 stratum radiatum are subject to \( \alpha 7 \) nAChR-mediated cholinergic modulation. It is unclear, however, whether the detailed distribution of \( \alpha 7 \) nAChRs at GABAergic synapses is similar to that at glutamatergic synapses.

**DISCUSSION**

Perhaps the most striking aspects of the current observations are the prevalence and density of \( \alpha 7 \) nAChR immunolabeling at synapses throughout the stratum radiatum. Almost all synaptic profiles in this region appear labeled over the synaptic membranes. Gold particles are also commonly found over presynaptic and postsynaptic elements of the synapse. The mean immunogold particle density at these \( \alpha 7 \) nAChR-labeled synapses (184.01 ± 10.15 particles/\( \mu m^2 \)) is remarkably close to the reported particle densities for NMDA and AMPA glutamate receptor labeling (≈200 particles/\( \mu m^2 \) for each) at synapses in rat CA1 stratum radiatum of similarly prepared tissue (Racca et al., 2000). The synaptic localization of \( \alpha 7 \) receptors reported here is consistent with immunogold labeling in guinea pig medial prefrontal cortex (Lubin et al., 1999); there also \( \alpha 7 \) nAChR-LIR was detected both presynaptically and postsynaptically, at axospinous (presumably glutamatergic) synapses and at a subset of double-labeled GABAergic synapses. The presence of \( \alpha 7 \) nAChR immunolabeling at presynaptic terminals, although consistent with reported nicotinic stimulation of hippocampal transmitter release (see below), is in contrast to the reported absence of terminal labeling as assessed by light microscopy (Domínguez del Toro et al., 1994).

The abundance of \( \alpha 7 \) nAChRs at these synapses raises questions about their physiological role. Functional \( \alpha 7 \) nAChR subunits have been demonstrated on hippocampal interneurons, where they constitute 38 pS, inwardly rectifying channels (Shao and Yakel, 2000) mediating strong excitatory effects (Jones and Yakel, 1997; Alkondon et al., 1998; Frazier et al., 1998a,b; McQuiston and Madison, 1999; Sudweeks and Yakel, 2000) including generation of action potentials. The \( \alpha 7 \)-mediated activation of such interneurons can result in either inhibition or disinhibition of pyramidal neurons (Ji and Dani, 2000). The presence of functional \( \alpha 7 \) subunits on hippocampal dentate granule or pyramidal cells has been more controversial. Activation of \( \alpha 7 \) nAChRs on mossy fiber presynaptic terminals has been found to increase intraterminal \( \text{Ca}^{2+} \) levels and to increase transmitter release (Gray et al., 1996), but others have failed to observe such \( \text{Ca}^{2+} \) elevation (Vogt and Regehr, 2001). Most investigators have found no \( \alpha 7 \) nAChR-mediated excitation of pyramidal neurons (Frazier et al., 1998b), despite the presence of very low but detectable levels of \( \alpha 7 \) nAChR subunit mRNA in these cells by single-cell RT-PCR (Sudweeks and Yakel, 2000). However, in acute and cultured hippocampal slices of 2- to 4-week-old rats, a small component of the EPSC evoked in CA1 pyramidal cells by extracellular...
stimulation in stratum radiatum has been reported to be α7 nAChR mediated (Hefft et al., 1999). The reasons for these different observations are not clear, but of some potential relevance may be the recent discovery of lynx1, an endogenous peptide homologous to αBgtx that enhances nicotinic receptor currents (Miwa et al., 1999). The distribution of lynx1 binding is similar to the distribution of α7 nAChRs, suggesting that physiological activation of α7 nAChRs may be modulated by the simultaneous binding of lynx1.

By producing inward, depolarizing current and particularly by directly mediating Ca\(^{2+}\) influx, activation of α7 nAChRs could be expected to influence synaptic transmission and plasticity. Presynaptic α7 nAChR activation, induced by repetitive, brief (5 × 200 msec at 8.5 sec intervals) application of 0.5 mM nicotine, can lead to persistent potentiation of glutamatergic synapses between dissociated hippocampal neurons (Radcliffe and Dani, 1998). α7 nAChR-mediated nicotinic activation in conjunction with postsynaptic depolarization has also been shown to induce LTP at glutamatergic synapses onto ventral tegmental area dopaminergic neurons. Evidence at those synapses suggested that activation of presynaptic α7 nAChRs most likely induced potentiation by acting presynaptically to increase the probability of glutamate release, and it was inferred that the consequent enhanced postsynaptic depolarization produced the NMDA receptor (NMDAR) activation necessary for LTP induction. There is also evidence that arachidonic acid, a putative retrograde messenger in LTP, can facilitate synaptic transmission by increasing α7 nAChR-mediated currents and thus enhancing presynaptic transmitter release (Nishizaki et al., 1999).

The abundance of postsynaptic α7 subunits revealed by our results also suggests, however, a possible postsynaptic locus for α7 nAChR-mediated synaptic potentiation. During early postnatal development, when AMPA receptors are absent from many postsynaptic membranes, the α7 nAChRs may actually mediate the induction of LTP, either directly or in conjunction with NMDARs. During this time, levels of α7 nAChRs in many brain areas, including hippocampus (Hunt and Schmidt, 1979) and somatosensory cortex (Bina et al., 1995), are much higher than in the adult. Activation of postsynaptic, particularly perisynaptic, α7 subunits, especially during the first postnatal week when their density is presumably highest, could depolarize a dendritic spine sufficiently to relieve the voltage-dependent Mg\(^{2+}\) block of simultaneously stimulated NMDA receptors, thereby inducing NMDA-dependent synaptic plasticity. Indeed, a selective α7 nAChR-mediated enhancement of the NMDA component of EPSPs recorded from auditory cortex pyramidal neurons is seen at postnatal day 8–16 but not in older rats (Aramakis and Metherate, 1998). The high Ca\(^{2+}\) permeability of α7 nAChRs (Bertrand et al., 1993; Seguela et al., 1993; Castro and Albuquerque, 1995) raises the further possibility that activation of these receptors alone could yield sufficient Ca\(^{2+}\) influx to trigger calcium-dependent processes, including the induction of synaptic plasticity (Ghosh and Greenberg, 1995). The likelihood of α7 nAChR-driven synaptic potentiation might be enhanced if the resulting spine depolarization were sufficient to activate voltage-gated calcium channels in the spine membrane (Yuste and Denk, 1995; Reid et al., 2001), or if the resulting Ca\(^{2+}\) influx were amplified by calcium-induced calcium release (CICR) from internal stores in the dendritic spine (Empetage et al., 1999); indeed, there is evidence for nicotinic activation of CICR (Empetage et al., 2001). Such mechanisms may not be essential for gross brain development, because transgenic mice lacking the α7 subunit displayed no abnormalities of brain anatomy (Orr-Urteger et al., 1997) or of behavior (Paylor et al., 1998). However, chronic systemic nicotine administration during the second week of postnatal development has recently been found to lead to persistent electrophysiological abnormalities in auditory neocortex (Aramakis et al., 2000). It is known that α7 nAChRs activate and desensitize rapidly in response to brief exposure to high ACh concentrations (Couturier et al., 1990); the half-maximal concentrations for α7 activation and inactivation of this subunit under physiological conditions appear to be in the range of 30–90 and 1–2 μM, respectively (Seguela et al., 1993; Fenster et al., 1997).

Choline, the product of ACh hydrolysis in the extracellular space, is a selective α7 nAChR agonist (Alkondon et al., 1997), and ambient levels of choline in the CSF caused by hydrolysis of ACh may be, after strong cholinergic activity, sufficiently high to activate and/or to desensitize α7 receptors (Papke et al., 1996). The half-maximal desensitization concentration is sufficiently high, however, for a large fraction of the receptors to remain functional under normal conditions (McGehee et al., 1995; Gray et al., 1996). The physiological consequence of these differential sensitivities is unclear, but it could provide a form of lateral inhibition in time and space: ACh release from an activated cholinergic terminal could result in α7-mediated facilitation of synaptic transmission and plasticity at simultaneously activated glutamatergic synapses.
synapses close to the activated terminal where the local ACh concentration would transiently be high. As a result of diffusion, synapses farther from the cholinergic terminal, or nearby synapses activated asynchronously, would experience inactivating ACh concentrations.

In addition to labeling of synaptic membranes, we also observed abundant labeling over endoplasmic reticulum and over membranous structures in the postsynaptic cytoplasm, including the spine apparatus (Figs. 5–7). This pattern resembles the reported association of β2 and α4 nAChRs with endoplasmic reticulum and transport vesicles in various neurons, including neocortical pyramidal cells (Hill et al., 1993; Nakayama et al., 1995). These observations of a large pool of intracellular α7 nAChRs suggest that these subunits may be actively internalized or inserted and that the extracellular, functional receptors may thus be dynamically regulated in response to the ongoing activity of the neuron.

Finally, the near-ubiquitous presence of α7 nAChRs at hippocampal synapses described here renders more salient the recent report (Wang et al., 2000b) of high-affinity binding of β-amyloid peptide (Aβ(1–42) to α7 nAChRs. Submicromolar concentrations of soluble Aβ(1–42), like its fibrillar amyloid precipitate, may be neurotoxic (Roher et al., 1996); toxicity can be partially blocked by nicotine (Wang et al., 2000a). Furthermore, similarly low concentrations of Aβ(1–42) can block α7 nAChR-mediated currents in hippocampal interneurons (Pettit et al., 2001). Thus the widespread distribution of α7 nAChRs in the hippocampus and their interactions with Aβ(1–42) peptide may be important factors in early cognitive impairments and later neuronal loss in Alzheimer’s disease.

REFERENCES


enhancement of fast excitatory synaptic transmission in CNS by pre-

McQuiston AR, Madison DV (1999) Nicotinic receptor activation ex-
cites distinct subtypes of interneurons in the rat hippocampus. J Neu-


Nakayama H, Shiota S, Okuda H, Nakashima T, Nakay (1995) Immuno-


Papke RL, Bencherif M, Lippiello P (1996) An evaluation of neuronal nicotinic acetylcholine receptor activation by quaternary nitrogen com-
ounds indicates that choline is selective for the alpha 7 subtype. Neurosci Lett 213:201–204.


Racca C, Stephenson FA, Streit P, Roberts JD, Somogyi P (2000) NMDA receptor content of synapses in stratum radiatum of the hip-


Radcliffe KA, Fisher JL, Gray R, Dani JA (1999) Nicotinic modulation of glutamate and GABA synaptic transmission of hippocampal neu-

Rapier C, Lunt GG, Wonnacott S (1988) Stereoselective nicotine-


Rocerig B, Nelson DA, Katz LC (1997) Fast synaptic signaling by nicoti-

Roher AE, Chaney MO, Kuo YM, Webster SD, Stine WB, Haverkamp LJ, Woods AS, Cotter RJ, Tuohy JM, Kraft GA, Bonnell BS, Em-


Segal M, Dudai Y, Amsterdam A (1978) Distribution of an α-bungaro-


Somogyi P, Takagi H (1982) A note on the use of picric acid-


Vidal C, Changeux J-P (1993) Nicotinic acid and muscarinic modula-

Vogt KE, Regehr WG (2001) Cholinergic modulation of excitatory syn-
aptic transmission in the CA3 area of the hippocampus. J Neurosci 21:75–83.


Wiedenmann B, Franke WW (1985) Identification and localization of synaptophysin, an integral membrane glycoprotein of MW 38,000 char-

Wonnacott S (1997) Presynaptic nicotinic ACh receptors. Trends Neu-
rosci 20:92–98.


Zarei MM, Radcliffe KA, Chen D, Patrick JW, Dani JA (1999) Distri-