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Building a Geochemical View of Microbial Salt Tolerance: Halophilic Adaptation of *Marinococcus* in a Natural Magnesium Sulfate Brine

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Current knowledge of life in hypersaline habitats is mostly limited to sodium and chloride-dominated environments. This narrow compositional window does not reflect the diversity of brine environments that exist naturally on Earth and other planetary bodies. Understanding the limits of the microbial biosphere and predicting extraterrestrial habitability demands a systematic effort to characterize ionic specificities of organisms from a representative range of saline habitats. Here, we investigated a strain of *Marinococcus* isolated from the magnesium and sulfate-dominated Basque Lakes (British Columbia, Canada). This organism was the sole isolate obtained after exposure to exceptionally high levels of Mg**2+** and SO**4**2− ions (2.369 and 2.840 M, respectively), and grew at extremes of ionic strength not normally encountered in Na**+**/Cl**−** brines (12.141 mol liter**−1**). Its association at the 16S rDNA level with bacterial halophiles suggests that ancestral halophily has allowed it to adapt to a different saline habitat. Growth was demonstrated in media dominated by NaCl, Na**2**SO**4**, MgCl**2**, and MgSO**4**, yet despite this plasticity the strain was still restricted; requiring either Na**+** or Cl**−** to maintain short doubling times. Water activity could not explain growth rate differences between media, demonstrating the importance of ionic composition for dictating microbial growth windows. A new framework for understanding growth in brines is required, that accounts for the geochemical history of brines as well as the various stresses that ions impose on microbes. Studies such as this are required to gain a truly universal understanding of the limits of biological ion tolerance.

Keywords: brine, habitability, *Marinococcus*, halophile, magnesium sulfate

INTRODUCTION

Hypersaline brines occur naturally on every continent on Earth, as well as on the ocean floor (Wallmann et al., 2002), in the subsurface (Hanor, 1994), and on other planetary bodies (Kargel, 2000; Tosca et al., 2008). Understanding the factors that limit and control microbial growth in brines is therefore crucial to charting the extent of the terrestrial biosphere, as well as predicting likely habitable environments off the Earth.

Knowledge of microbial life in saline systems is largely based on observations of sodium and chloride-dominated environments, which form the majority of hypersaline water bodies on Earth.
between generic salt tolerance and specific adaptation to Na+ ‘halophilic’ and ‘halotolerant,’ as often no distinction is made as ion-specific? Can we apply generally to all brine environments? And how much shown that high ionic strength brines rich in divalent ions can powerfully limits microbial growth (Hallsworth et al., 2003; Madern et al., 2000). Many of these historical studies, whilst they suggest important ion-specific effects, do not account for variations in water activity across different salt solutions, thus making discrimination between ion-specific and hydration-related effects difficult.

More recently, observations of deep-sea hypersaline anoxic basins that contain magnesium chloride brines have shown that chaotropicity (macromolecule-disordering effects), not water activity, defines microbial colonization in the brine: seawater interface (Hallsworth et al., 2007; Yakimov et al., 2015). Chaotropicity is a property of ion combinations, such as Mg²⁺/Cl⁻, that is not exhibited by Na⁺/Cl⁻ brines and which powerfully limits microbial growth (Hallsworth et al., 2003; Ball and Hallsworth, 2015). Furthermore, a previous study has shown that high ionic strength brines rich in divalent ions can limit microbial growth even when water activity is sufficiently permissive (Fox-Powell et al., 2016). Observations such as these raise the questions: how much of what has been learnt about microbial salt adaptation from studying Na⁺/Cl⁻ environments can we apply generally to all brine environments? And how much is ion-specific?

It is currently unclear whether a tendency for generic salt tolerance exists. There is even uncertainty surrounding the words ‘halophilic’ and ‘halotolerant,’ as often no distinction is made between generic salt tolerance and specific adaptation to Na⁺ and Cl⁻. Here, we use the prefix ‘halo’ when discussing specific Na⁺ and Cl⁻ adaptation. Some authors have used similar terms to describe specific tolerance of other salts (e.g., ‘epsophily’ for life in high Mg²⁺/SO₄²⁻ environments; Crisler et al., 2012, or ‘natronophil’y for life in high Na⁺/HCO₃⁻ environments; Banciu and Sorokin, 2013), but such terms are not employed here. The significance of these distinctions (including the widely used ‘halophile’) will remain unclear until it is understood how the effects of different ions manifest themselves on an organismal scale, particularly as most brine environments are characterized by dynamic assemblages of salt ions.

A necessary first step in this effort is to systematically study organisms from a diverse range of hypersaline environments, to understand their ion specificities both as a function of and independent from water activity. If halophilic tendencies equip organisms to inhabit non-Na⁺/Cl⁻ brines, then it can be expected that inhabitants of such environments will themselves be halophiles or descendants of halophiles. Alternatively, if halophilic adaptation does not permit growth in other saline environments, it should be expected that brine inhabitants are descended from a variety of clades, regardless of ancestral halophily. Molecular surveys of environments such as the Dead Sea, which contains high concentrations of Mg²⁺ alongside Na⁺ and Cl⁻, have revealed dynamic populations of halophiles whose structures fluctuate in response to changes in ionic composition (Bodaker et al., 2010), whilst the Mg²⁺/SO₄²⁻-rich Spotted Lake (British Columbia, Canada), and Hot Lake (Washington, United States) notably lack the halophilic archaea, which are ubiquitous inhabitants of Na⁺/Cl⁻ brines (Lindemann et al., 2013; Pontefract et al., 2017). Laboratory-produced sulfate brines have also been shown to select for communities lacking typical halophiles, even when such organisms were present in the inoculum (Fox-Powell et al., 2016). Developing a truly universal understanding of the limits of biological ion tolerance requires marrying such community-level studies with in-depth investigations of the physiological limitations and requirements of specific microorganisms.

Here, we used the Basque Lakes, British Columbia, Canada as a natural laboratory to explore biological adaptation to non-Na⁺/Cl⁻ brines. Like neighboring Spotted Lake and Hot Lake, these are a series of Mg²⁺/SO₄²⁻ dominated hypersaline lakes that in the summer are concentrated to beyond epsomite (MgSO₄ 7H₂O) saturation (Eugster and Hardie, 1978). They bear one of the highest reported divalent: monovalent ion ratios in natural brines and as such are an important analog for divalent ion-rich brines on Mars and elsewhere in the solar system (Foster et al., 2010; Fox-Powell et al., 2016). We report the isolation of a strain of Marinococcus, which was capable of growth in extremely high concentrations of Mg²⁺ and SO₄²⁻. The clustering of this genus with other typical halophiles suggests that the ancestral halophily of Marinococcus strain IS₁₈B₂ has allowed it to adapt to grow under high magnesium sulfate conditions.

**MATERIALS AND METHODS**

**Field Sampling and Geochemical Analyses**

The Basque Lakes are a series of magnesite and sulfate brine lakes, pools and playas in the semi-arid interior of British Columbia, Canada (Figure 1). The lakes experience ion concentrations beyond epsomite saturation in the dry summer months (Eugster and Hardie, 1978). Basque Lakes no. 1 (N 50° 36.01 W 121° 21.31) and 2 (N 50° 35.35 W 121° 20.54) were sampled in February 2015. The lakes consisted of numerous smaller ice-covered pools (Figure 1). Brine-inundated sediments were sampled from three pools (one from Lake no. 1 and two from Lake no. 2) using sterile 50 ml tubes and spatulas sterilized.
in 70% ethanol. Tubes were filled to the top to minimize gas phase, sealed with parafilm and kept at 4°C until shipment back to the laboratory. Oxidation/reduction potential, pH and temperature were measured in situ using a Myron L Company Ultrameter II and a Fisher Scientific Pt-100 platinum sensor digital thermometer. Lake waters for geochemical analyses were filtered (0.22 µm) and stored in 15 ml tubes for shipment back to the laboratory. Samples intended for cation analysis were acidified with HNO₃ to a final concentration of 1%.

Major cations were analyzed by Inductively Coupled Plasma Optical Emission Spectroscopy (ICP-OES) at The University of Edinburgh, according to manufacturer’s instructions. Major anions were analyzed by Ion Chromatography (IC) at The University of Edinburgh, in both forward and reverse directions. The University of Edinburgh, United Kingdom, using a Dionex DX-120 system according to the manufacturer’s instructions.

**DNA Extraction, Sequencing, and Phylogenetic Analysis**

Strain IS₅B2c was grown to stationary phase in BL medium, harvested by filtration onto sterile 0.22 µm-pore 25 mm diameter polycarbonate filters (Merck Millipore) and DNA was extracted using a modified phenol:chloroform:isoamyl alcohol protocol as described in Urakawa et al. (2010). DNA was interrogated by polymerase chain reaction (PCR) using the primers 27F (5'-AGAGTTTGATCMTGGCTCAG-3') and 1389R (5'-ACGGGCGGTGTGAGG-3') which target bacterial 16S ribosomal RNA (Cameron et al., 2012). Each individual 25 µl PCR reaction contained 1 µl template, 0.4 µM of the relevant forward and reverse primer, 200 µM dNTPs, 1.5 mM MgCl₂, 1× PCR buffer and 1 unit Taq polymerase (Inveritox). Reactions were treated at 95°C for 5 min and then subject to 25 cycles of: melting at 95°C for 30 s, annealing at 55°C for 30 s and extension at 72°C for 30 s, and finished with a final extension step at 72°C for 10 min. Bacterial PCR products were purified using the Qiagen Amplicon Purification Kit according to manufacturer’s instructions, and sequenced on the Sanger ABI 3730 XL platform at Edinburgh Genomics, The University of Edinburgh, in both forward and reverse directions.
Sequence processing, trimming and contig assembly was performed using the DNA Baser software. First, primer sequences were removed from both complementary sequences. Next, the sequences were aligned for contig assembly and low-quality ends and ambiguous bases were removed. The final consensus sequence was exported for phylogenetic analysis and aligned with the NCBI database using the BLASTn tool. Phylogenetic analysis was performed using the CLUSTAL_W program in the MEGA v. 7 software suite. A phylogenetic tree was constructed using the neighbor joining approach (Saitou and Nei, 1987), with Alicyclobacillus acidocaldarius used as the outgroup (Wang et al., 2009). The topology of the tree was evaluated by the bootstrap resampling method based on 1000 replicates.

**Growth Response Quantification**

Growth of strain IS$_{B2c}$ was quantified by measuring increase in optical density at 600 nm ($OD_{600}$) in 96 well plates in a Biotek Synergy 2 microplate reader. Doubling times were calculated from the exponential growth portion of $OD_{600}$ curves. The pH-defined growth range of strain IS$_{B2c}$ was investigated by recording $OD_{600}$ increase in a series of BL media buffered in 0.5 pH unit increments between pH 3.0 and pH 9.0. Levels of pH above 7.5 were achieved by the addition of Tris; levels below pH 6.5 were adjusted with HCl. Temperature ranges were established by recording $OD_{600}$ increase in BL medium (pH 7.0) at increments of increasing temperature from 4 to 45°C. The NaCl requirement (halophily) of strain IS$_{B2c}$ was established in a series of NaCl media, in which the most concentrated contained (per liter) 292.2 g/5.0 M NaCl and concentration was decreased systematically comparing growth rates between media that contained 0.746 g/0.010 M KCl, 4 g yeast extract (Oxoid) and 3 g casamino acids (Difco). Apart from the main salt dissolved, the media contained (per liter) 0.746 g/0.010 M KCl, 4 g yeast extract (Oxoid) and 3 g casamino acids (Difco).

The ionic requirements of strain IS$_{B2c}$ were tested by systematically comparing growth rates between media that contained 2 M of each of four ions (Mg$^{2+}$, Na$^{+}$, SO$_4^{2-}$, and Cl$^{-}$) in all possible cation-anion pairings. Apart from the main dissolved salt, the media contained (per liter) 0.746 g/0.010 M KCl, 4 g yeast extract (Oxoid) and 3 g casamino acids (Difco).

**Water Activity Measurement**

Water activity was quantified at 30°C using a Rotronic HP23-AW water activity meter in AWE mode, calibrated to five points (0.325, 0.595, 0.755, 0.845, and 0.935) using saturated salt solution standards (MgCl$_2$, NH$_4$NO$_3$, NaCl, KCl, and KH$_2$PO$_4$, respectively) prepared as described by Winston and Bates (1960).

**RESULTS**

**The Basque Lakes**

At time of sampling, the waters of both Basque Lake no. 1 and Basque Lake no. 2 were in a relatively dilute phase likely due to increased rain and snowmelt input and lower evaporation rates in winter. The three sampled pools (distributed between Lakes nos. 1 and 2) possessed ionic strengths between 0.498 and 1.038 mol liter$^{-1}$, magnesium/sodium ratios between 1.50 and 3.56 and sulfate/chloride ratios between 28.31 and 86.92 (Table 1).  

**Marinococcus Strain IS$_{B2c}$**

A single isolate, originating from Basque Lake no. 2 and designated strain IS$_{B2c}$, was obtained by serial dilution-to-extinction. Growth in the high ionic strength (12.141 mol liter$^{-1}$) enrichment medium was slow; taking around 14 days for cell numbers to increase from inoculum values in microscopic observations. Cells of strain IS$_{B2c}$ were 1–2 μm diameter coccoids (Figure 2), which when grown to high density in the routine BL medium exhibited a noticeably visible orange-yellow pigmentation.

A partial 16S rDNA sequence was obtained, covering 1,117 base pairs. At the 16S rDNA level, strain IS$_{B2c}$ clustered most closely with the moderately halophilic, aerobic Gram-positive bacterial genus *Marinococcus*, sharing >99% identity with all other described species in the genus (Figure 3). BLAST analysis revealed the closest database match at 16S rDNA level to be from a bacterial culture originating from Sehline Sebkha salt lake, Tunisia, which also clustered in the *Marinococcus* genus (Hedi et al., 2014).

Similar to previously described *Marinococcus* strains, strain IS$_{B2c}$ grew optimally at pH 7.0, with short doubling times also recorded at pH 6.5 and 7.5. No growth was observed below pH 6.0 or above pH 7.5 (Figure 4). Strain IS$_{B2c}$ grew optimally at a similar temperature to other *Marinococcus* isolates, which have temperature optima around 30°C (Novitsky and Kushner, 1976; Van Hao et al., 1984; Li et al., 2005; Wang et al., 2009) and was capable of slow growth at 15°C (growth rate not quantified), but not at 4°C, or at 45°C (Figure 4).

**Growth of Strain IS$_{B2c}$ in Varied Ionic Regimes**

NaCl requirements for strain IS$_{B2c}$ were similar to other members of the genus; growth was demonstrated over a NaCl concentration range of 0.5–4.0 M (approximately 3–24%), with the fastest growth rate around 1.75–2 M (Figure 5). No growth

<table>
<thead>
<tr>
<th>Basque 1</th>
<th>Basque 2b</th>
<th>Basque 2c</th>
</tr>
</thead>
<tbody>
<tr>
<td>K$^+$</td>
<td>0.094</td>
<td>0.184</td>
</tr>
<tr>
<td>Na$^+$</td>
<td>0.645</td>
<td>1.543</td>
</tr>
<tr>
<td>Mg$^{2+}$</td>
<td>2.105</td>
<td>2.449</td>
</tr>
<tr>
<td>Ca$^{2+}$</td>
<td>0.172</td>
<td>0.196</td>
</tr>
<tr>
<td>SO$_4^{2-}$</td>
<td>9.051</td>
<td>11.950</td>
</tr>
<tr>
<td>Cl$^{-}$</td>
<td>0.100</td>
<td>0.156</td>
</tr>
<tr>
<td>Mg: Na</td>
<td>3.084</td>
<td>1.502</td>
</tr>
<tr>
<td>SO$_4$:Cl</td>
<td>33.301</td>
<td>28.306</td>
</tr>
<tr>
<td>Ionic strength/ mol liter$^{-1}$</td>
<td>0.387</td>
<td>0.498</td>
</tr>
<tr>
<td>Temperature/°C</td>
<td>0.9</td>
<td>0.9</td>
</tr>
<tr>
<td>pH</td>
<td>6.23</td>
<td>6.26</td>
</tr>
<tr>
<td>Conductivity/mS</td>
<td>13.9</td>
<td>28.6</td>
</tr>
<tr>
<td>ORP (mV)</td>
<td>−150</td>
<td>nd</td>
</tr>
</tbody>
</table>

**Table 1 | Ionic composition and physicochemical parameters of the Basque Lakes sample sites.**
was observed at 5 M or at 0.25 M NaCl. The doubling time of *Marinococcus* strain IS$_5$B2c was approximately the same in BL medium (dominated by Mg$^{2+}$ and SO$_4^{2-}$) as in the optimum concentration of pure NaCl.

Strain IS$_5$B2c showed remarkable plasticity with its ionic requirements, growing in media where the main salts were NaCl, Na$_2$SO$_4$, MgCl$_2$, or MgSO$_4$ (Figure 5). Growth rates across the different media compositions varied considerably. The most rapid growth was measured in 2 M Na$^+/\text{Cl}^-$, and the slowest in 2 M Mg$^{2+}$/SO$_4^{2-}$.2 M SO$_4^{2-}$ (paired with Na$^+$) supported a comparable growth rate to 2 M Cl$^-$(paired with Mg$^{2+}$). Doubling times in 2 M Na$^+$ when paired with SO$_4^{2-}$ were slightly longer (Figure 5).

To attempt to explain the different growth rates, we measured water activity in all growth media. We found that when grown in NaCl, strain IS$_5$B2c had a water activity optimum of approximately 0.940 $a_w$ with similar growth rates obtained between 0.950 $a_w$ and 0.920 $a_w$ (Figure 6). Higher and lower water activities produced progressively longer doubling times. Synthetic Basque Lakes medium (BL medium), at 0.925 $a_w$, supported rapid doubling times in agreement with the optimum water activity range in NaCl. Water activity varied considerably across the other salt media, despite sharing absolute concentrations of specific ions. Most ion pairings produced doubling times that fell on or near the NaCl water activity growth response curve, with the notable exception of 2 M Mg$^{2+}$/Cl$^-$ (which prohibited all growth) and 2 M Mg$^{2+}$/SO$_4^{2-}$, which had a water activity near the optimum value yet a growth rate close to the slowest recorded here (Figure 6).

**DISCUSSION**

The Basque Lakes

The Basque Lakes, British Columbia are part of a series of hypersaline playas in western North America that exhibit unusual chemistries (Renaut, 1990). The Basque Lakes are particularly unique for not simply containing Mg$^{2+}$ and SO$_4^{2-}$ as major ions, but for being dominated by them, reflected in high Mg$^{2+}$/Na$^+$ ratios and even higher SO$_4^{2-}$/Cl$^-$ ratios (Table 1). This water chemistry is similar to that found in nearby Spotted Lake (Pontefract et al., 2017). Such environments provide valuable natural laboratories for understanding microbial salt tolerance, and due to their high divalent : monovalent ion ratios make compelling analogs for sulfate brines on ancient Mars, which likely possessed divalent ion content highly atypical of terrestrial chloride brines (Vaniman et al., 2004; Tosca et al., 2011). Indeed, sulfates (Mg, Fe, and Ca) make up the majority of martian salt minerals both in evaporate outcrops (Bibring et al., 2006; Hynek et al., 2015), and in the globally distributed dust (Yen et al., 2005), recording the widespread presence of sulfate-rich brines during a key transitional phase of martian surface evolution. It follows that properly understanding how martian habitability has changed through time requires knowledge of how life adapts to cope with highly saline sulfate-rich environments. Thus, isolates from environments such as the Basque Lakes can help inform what establishes boundaries to biologically permissible environments on other planetary bodies, whether that be indigenous life or less speculatively, microorganisms transferred into these environments by robotic or human missions.

**Marinococcus Strain IS$_5$B2c: A Magnesium and Sulfate-Loving Halophile**

A strain of *Marinococcus*, designated IS$_5$B2c, was isolated from the Basque Lakes using a high ionic strength growth medium. The successful cultivation of strain IS$_5$B2c at an ionic strength of 12.141 mol liter$^{-1}$ in IS$_5$ medium, with a Mg$^{2+}$ concentration of 2.369 M and a SO$_4^{2-}$ concentration of 2.840 M represents both the highest documented ionic strength and the highest co-occurring concentrations of Mg$^{2+}$ and SO$_4^{2-}$ directly demonstrated to be tolerated by a microorganism. Growth at these levels is remarkable, particularly considering that high ionic strength (>10 mol liter$^{-1}$) can interact with other stressors to powerfully limit microbial growth, precluding halophiles that tolerate more extreme levels of low water activity (Fox-Powell et al., 2016).

Similar growth rates recorded in BL medium (dominated by Mg$^{2+}$ and SO$_4^{2-}$ with Na$^+$ and Cl$^-$ approximately an order of magnitude lower in concentration) and in the optimum concentration of pure NaCl (Figure 5) is a likely sign of adaptation to its source environment. The replacement of Na$^+$/Cl$^-$ with Mg$^{2+}$/SO$_4^{2-}$ as the dominant ions with no negative effect on growth hints at a set of evolutionary adaptations in the Basque Lakes which could supply a host of valuable information on the ion specificity of salt stress.

The *Marinococcus* genus is classified at the high end of moderate halophily; members typically grow optimally over the range 10–15% dissolved NaCl (approximately 1.8–2.6 M), but some can tolerate higher concentrations (up to 25%) (Li et al., 2005; Rice et al., 2009). The placing of strain IS$_5$B2c within the *Marinococcus* genus, and this genus's close phylogenetic
association with other halophilic genera (including *Pontibacillus*, *Halobacillus*, and *Salimicrobium*) is a clear demonstration that halophilic (NaCl-loving) organisms are not precluded from hypersaline environments of fundamentally different compositions. It is likely that physicochemical stressors common to all hypersaline environments, such as low water activity, ensure that halophilic organisms already equipped to deal with them have a competitive advantage over those that are not.

However, other strains of *Marinococcus* are associated with brine compositions that suggest deviations from traditional halophily. The closest database match for strain IS$_5$B2c was an isolate from Sehline Sebkha salt lake, Tunisia, which is dominated by Na$^+$ and Cl$^-$ but with significant concentrations of Mg$^{2+}$ and SO$_4^{2-}$ as secondary ions (Hedi et al., 2014). *Marinococcus halotolerans*, isolated from hypersaline soils in China, was initially enriched in a medium containing 250 g L$^{-1}$ MgCl$_2$.6H$_2$O (1.23 M MgCl$_2$), with relatively low concentrations of NaCl (2 g L$^{-1}$; 33 mM) (Li et al., 2005). The variety of saline conditions tolerated by this organism are remarkable; the authors report similar growth responses across wide concentration ranges of NaCl, KCl, and MgCl$_2$, which is rare for halophilic or halotolerant organisms which usually require NaCl to some degree (Boring et al., 1963; Javor, 1984; Ventosa et al., 1998; Crisler et al., 2012). This plasticity is also reflected in strain IS$_5$B2c, as discussed below. Enrichments done in high levels of Mg$^{2+}$ by Li et al. (2005) and in the current study both resulted in the culture of a strain of *Marinococcus*, despite the large degree of geographical separation between the source environments. Due to their ability to tolerate a seeming wide variety of ionic compositions, including those of extreme ionic strength, species within the *Marinococcus* genus make compelling model organisms for understanding the capacity for life in hypersaline environments that differ significantly from typical Na$^+$/Cl$^-$ brines.

**Beyond Water Activity: Ion Specificities of *Marinococcus* Strain IS$_5$B2c**

Developing a universal understanding of microbial ion tolerance requires systematic assessments of the ionic requirements of organisms from a range of hypersaline environments. It is only with an appreciation of how ionic composition defines growth in the context of an organism’s phylogenetic associations that we can begin to discriminate between generic salt adaptation tendencies and ion-specific effects. Historical efforts to investigate the ionic dependencies of halophiles produced results that frustrated attempts to generalize; organism-specific idiosyncrasies seemed to be the rule (e.g., Javor, 1984; Ventosa et al., 1998). Consistencies that did emerge were that all halophiles require Na$^+$ to some degree, and that the requirement for Na$^+$ and Cl$^-$ ions could be lowered somewhat through substitution with other solutes (including organic solutes such as glucose or glycerol) (Adams et al., 1987). However, these studies lacked systematic consideration of other powerful growth-mediating physicochemical parameters, such as water activity, which can vary significantly across different brines.

Water activity is a measure of the thermodynamic availability of water, expressed relative to pure water at the same temperature and pressure. The degree to which water activity is depressed in an aqueous system varies with the solute. For example, 2 M solutions of NaCl and MgSO$_4$ have respective water activities of approximately 0.93 $a_w$ and 0.96 $a_w$, which is greater than the difference between freshwater and seawater. Given that

![FIGURE 3](https://example.com/figure3.png)
water activity is a powerful mediator of microbial growth (Stevenson et al., 2015), any assessment of the effects of ionic composition on microbial growth requires consideration of this parameter.

Strain IS$_5$B$_2$c was therefore subjected to suite of growth trials in which the main dissolved salt was varied systematically (by concentration), and water activity was measured across all media. When strain IS$_5$B$_2$c was grown in different pure salt media, growth response varied considerably, despite the media sharing absolute concentrations of ions; as has been observed in many strains (Abram and Gibbons, 1961; Boring et al., 1963; Javor, 1984; Ventosa et al., 1998). It is tempting to interpret this as preferences for certain ions; for example in both 2 M Cl$^-$ and 2 M SO$_4^{2-}$, doubling times were shorter if the cation was Na$^+$ than Mg$^{2+}$ (Figure 6), inviting the conclusion that strain IS$_5$B$_2$c has a preference for Na$^+$ over Mg$^{2+}$.

However, considering water activity alongside ionic composition explained some of strain IS$_5$B$_2$c’s apparent ionic preferences. An apparent preference for Na$^+$ over Mg$^{2+}$ when paired with 2 M Cl$^-$ was satisfactorily explained by a higher water activity in the Mg$^{2+}$/Cl$^-$ media (which had a concentration of 1 M); meaning that it lay just outside the organism’s optimum range (Figure 6). When the 2 M Cl$^-$ (with Mg$^{2+}$) growth rates were compared with NaCl media at approximately the same water activity, doubling times were in excellent agreement despite differences in absolute Cl$^-$ concentration (Figure 6). This is a clear demonstration that if ionic preferences are to be understood, physicochemical factors such as water activity must be accounted for.

Water activity alone cannot be used to predict doubling time, however, otherwise all doubling times would have fallen somewhere on the optimum curve, which was not the case. Clearly, ion-specific effects were important. 2 M Mg$^{2+}$ paired with Cl$^-$ inhibited all growth, despite possessing a water activity that was demonstrably permissive in NaCl. This is likely due to the highly chaotropic activity of MgCl$_2$ at these concentrations.
by high ionic strength or some toxic effect of either Mg
levels of these ions. Again, this cannot be explained
than that predicted by the water activity curve in NaCl (Figure 6).

4 higher than the pure MgSO$_4$ recorded in 2 M MgSO$_4$
dilute conditions.

doubling times slightly faster than that predicted from water
function of water activity. Pure salt media (including all possible cation-anion
pairings) are the same as those reported in Figure 5. Green shaded area
represents the water activity response curve (±1 standard error) of the strain
when grown in pure NaCl media. Square symbols indicate sulfate salts,
triangles indicate chloride salts. Filled symbols indicate media containing
either 2 M of the anion or 2 M of both anion and cation (in the case of salts
with a 1:1 stoichiometry). Empty symbols indicate media containing 2 M of
the cation only. BL = BL medium, plotted for reference. Note that 2 M Mg$^{2+}$
paired with chloride inhibited all growth (inverted red triangle). Error bars
represent ±1 standard error of triplicate experiments.

It has previously been shown that chaotropic solutes such as MgCl$_2$
can impose stresses on microorganisms even when water activity is at permissive levels
(Hallsworth et al., 2003), and currently biological activity of any
kind above 2.3 M MgCl$_2$ has not been documented (Hallsworth et al., 2007; Yakimov et al., 2015). Both Na$_2$SO$_4$ media produced
doubling times slightly faster than that predicted from water
activity alone, suggesting a preference for sulfate under more
dilute conditions.

Most striking are the notably long doubling times
recorded in 2 M MgSO$_4$, indicating a much slower growth rate
than that predicted by the water activity curve in NaCl (Figure 6).
This is despite being isolated from an environment characterized
by high levels of these ions. Again, this cannot be explained
by high ionic strength or some toxic effect of either Mg$^{2+}$ or
SO$_4^{2-}$, as the strain grew readily in BL medium (dominated
by Mg$^{2+}$ and SO$_4^{2-}$ with an ionic strength ~0.5 mol liter$^{-1}$
higher than the pure MgSO$_4$ medium), at the shortest doubling
time achieved for the strain. It is important to note, however,
that BL medium also contained Na$^+$ and Cl$^-$ as secondary ions.

When both Na$^+$ and Cl$^-$ were omitted from the growth
media, growth was dramatically retarded. Surprisingly, it was
not a requirement for Na$^+$ and Cl$^-$ in combination that defined
this limitation, as the organism would grow well in media where
one of either Cl$^-$ or Na$^+$ had been removed (for instance, in
1 M MgCl$_2$ or 1 M Na$_2$SO$_4$). Rather, if either of these
ions were present in appreciable quantities, it was sufficient to
stimulate growth. This is remarkable, as cations and anions are
handled by wholly different cellular pathways and thus it is highly
implausible that there could be some active cellular process that
could readily substitute Na$^+$ for Cl$^-$. It could instead be due to
some affinity for specific ions in the structure of the organism’s
biomolecules, as is observed in the halophilic archaea, and also in
the membrane proteins of some bacterial halophiles (Oren et al.,
2005).

The Haloarchaea have evolved to accept molar quantities of
K$^+$ (in place of Na$^+$) and Cl$^-$ into the cell, and their intracellular
machinery must be structurally modified to function in the
presence of these ions (Soppa, 2006). The observed changes
mostly decrease hydrophobicity and increase protein affinity
for water molecules, both of which seem to be approaches to
mitigate low water activity stress (Tadeo et al., 2009). However,
enzymes from these organisms are shown to have ion specificities
beyond a simple need for hydration, and function optimally only
if K$^+$ and Cl$^-$ are provided (Ortega et al., 2011; Karan et al.,
2012). A narrow window of ion-specificity in the Haloarchaea
might explain their absence from Mg$^{2+}$/SO$_4^{2-}$ environments
like Spotted Lake, British Columbia (Pontefract et al., 2017),
and sulfate brine enrichments (Fox-Powell et al., 2016), as
well as the loss of Haloarchaeal dominance in the Dead Sea
following evaporative concentration of Mg$^{2+}$ (Bodaker et al.,
2010).

Due to an ostensible lack of these proteome modifications
in halophiles that maintain low cytoplasmic ion concentrations,
such as most halophilic bacteria (including the *Marinococcus*
genus; Louis and Galinski, 1997), it could be predicted that
such organisms are more plastic with their ionic requirements,
responding predominately to changes in water activity. However,
studies have shown that even bacterial halophiles still bear some
of the signatures of ‘salt-in’ Haloarchaeal proteomes, particularly
in their membrane bound proteins which are exposed to the
full salinity of the external milieu (Oren et al., 2005). Some
specificity should therefore be expected, even in halophiles whose
cytoplasmic salt ion content never rises beyond that of non-
halophiles. Our data support this prediction, as growth appeared
to be controlled predominately by hydration (water activity)
effects as long as either Na$^+$ or Cl$^-$ were available; when both
of these ions were removed, growth was dramatically slowed
(Figure 6).

Future work should focus on determining the mechanisms
employed by strain IS$_5$B2c for coping with extremely high
Mg$^{2+}$ and SO$_4^{2-}$ concentrations. In particular, changes in
metabolic pathways and substrate utilization in strain IS$_5$B2c
under different saline conditions is important follow-on work. As
Mg$^{2+}$ concentrations in the Dead Sea became elevated, archaeal
CorA magnesium channels became enriched 11-fold (Bodaker
et al., 2010). It was assumed this was a resistance mechanism
in the halophilic archaea to cope with the changing composition,
and if so implies that the typically ‘salt-in’ Haloarchaea were
actively excluding the divalent cation. If this energy-demanding
process is the only approach available to organisms in high Mg$^{2+}$
environments then that may well be a determining factor for
brine habitability.
Toward a Geochemical View of Salt Ion Tolerance: Lessons Learned From a Model Strain

Most knowledge of how microorganisms grow in hypersaline environments is drawn from studies of sodium and chloride-rich brines. Whilst this approach has served us well in understanding the distribution and activity of halophiles in the most dominant brine type on Earth, it has limited our ability to predict the extent of microbial habitat into other brine types, including those that exist in evaporites in the deep subsurface (e.g., Payler et al., unpublished) or on other planetary bodies (Fox-Powell et al., 2016).

A future goal of mapping the limits of life in high salt is to be able to analyze the composition of any brine and make informed, accurate predictions about habitability. Trade-offs almost certainly exist, as coping with high levels of salts is energetically expensive, meaning that certain ionic compositions might preclude low-energy metabolisms (Oren, 2011). Additionally, if protein structural changes always bear ion specificity, adaptations required to cope with one major ion might always decrease tolerance of another. These questions and others can be answered with systematic efforts to characterize further organisms from diverse brine types. It is notable that even in a brine fundamentally different from typical Na\(^+\)/Cl\(^-\) environments, halophiles such as Marinococcus can thrive, demonstrating that halophily does bestow aspects of a generic salt tolerance. The predictive power of water activity for growth rate was demonstrated across some pure salts, suggesting a limited ionic plasticity of this form of halophily. However, ionic specificity remained; in the absence of Na\(^+\) and Cl\(^-\), water activity no longer satisfactorily predicted growth rate.

This finding raises many questions about what might be required on a molecular level to develop true independence from sodium and/or chloride, and whether this has occurred on Earth. It is an open question whether organisms from hypersaline environments can be fully composition-independent and respond only to water activity, or if some ion-specificity will always occur. If this requirement for either Na\(^+\) or Cl\(^-\) is indeed a residual trait from strain IS5B2c’s halophilic ancestors, would inhabitants of Mg\(^{2+}\)/SO\(_4^{2-}\) brines that don’t share this evolutionary history exhibit a different set of requirements? Such organisms, should they exist, should be the target of future cultivation efforts.

AUTHOR CONTRIBUTIONS

CC and MF-P obtained the samples and contributed equally to experimental design and interpretation of results. MF-P conducted laboratory work and wrote the manuscript. CC contributed text, reviewed and edited the manuscript.

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