



Open Research Online

The Open University's repository of research publications and other research outputs

Modelling the injured spinal cord using 3-dimensional cell cultures; strategies for improving tissue engineered repair

Journal Article

How to cite:

East, E.; Golding, J. P. and Phillips, J. B. (2008). Modelling the injured spinal cord using 3-dimensional cell cultures; strategies for improving tissue engineered repair. *European Cells and Materials*, 16(Supple), p. 51.

For guidance on citations see [FAQs](#).

© 2008 AO Foundation

Version: Version of Record

Link(s) to article on publisher's website:

<http://www.ecmjournal.org/journal/supplements/vol016supp03/pdf/v016supp03a051.pdf>

Copyright and Moral Rights for the articles on this site are retained by the individual authors and/or other copyright owners. For more information on Open Research Online's data [policy](#) on reuse of materials please consult the policies page.

Modelling the injured spinal cord using 3-dimensional cell cultures; strategies for improving tissue engineered repair

E East, J P Golding & J B Phillips

Department of Life Science, The Open University, Walton Hall, Milton Keynes, UK.

INTRODUCTION: Traumatic injuries to the spinal cord are debilitating and often lead to paralysis and loss of sensation. CNS injury results in the formation of a glial scar that is largely composed of reactive, hypertrophic astrocytes which upregulate various markers including glial fibrillary acidic protein (GFAP) and sulphated proteoglycans. The glial scar microenvironment is inhibitory for axon growth and regeneration. A common finding of strategies aimed at bridging CNS lesions [1], in particular recent tissue engineered approaches using fibronectin [2], is that whilst axons readily enter and traverse the bridging graft they generally fail to exit the graft and re-enter the parenchyma due to the inhibitory glial scar at the graft-CNS interface. The aim of this work was to create a 3D cell culture model of CNS reactive gliosis, in which improved tissue-engineered repair strategies can be developed.

METHODS: Primary rat astrocytes were seeded onto collagen coated coverslips (2D) or into 3D collagen gels and the expression of markers associated with glial scar formation were analysed by immunocytochemistry (GFAP, chondroitin sulphate proteoglycan (CSPG), vimentin, aquaporin 4 (AQP4) and S100 β). To stimulate reactive gliosis, 3D astrocyte cultures were treated with TGF β 1 (10ng/ml) every other day for 15 days [3]. Cultures without TGF β 1 were used as controls. Expression of glial scar markers was assessed and measured using Volocity image analysis software (Improvision). Cell perimeter was measured as an indicator of astrocyte activation. Area of immunoreactivity was measured for CSPGs released from astrocytes.

RESULTS: Astrocytes in 3D cultures had significantly less immunoreactivity for GFAP, CSPG, vimentin, AQP4 and S100 β , than astrocytes in 2D cultures indicating a lower level of activation in 3D (fig 1). However immunoreactivity for β -actin did not differ in 2D or 3D. Following treatment with TGF β 1, astrocytes in 3D became hypertrophic and reactive, with changes in the expression of gliosis markers over time relative to control cultures (fig 2).

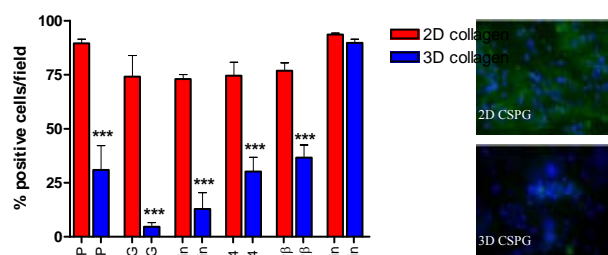


Fig. 1. Comparison of astrocyte activation markers in 2D and 3D cultures in type I collagen. Bars represent mean \pm SD, *** $P < 0.001$ 2D vs 3D.

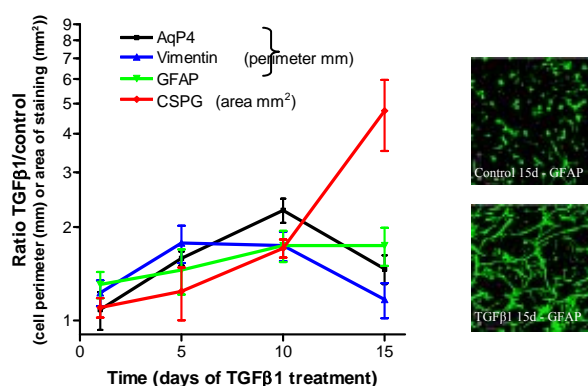


Fig 2. Increased astrocyte activation following TGF β 1 treatment of 3D cultures.

DISCUSSION & CONCLUSIONS: Astrocytes in 3D are less reactive than their 2D counterparts, thus resembling the physiological *in vivo* situation and providing a useful system in which activation can be investigated. TGF β 1 treated astrocytes over 15 days take on characteristics of the mature glial scar, including their morphology and protein expression profile. This model will provide a useful test-bed for tissue engineered strategies to improve axonal growth through the glial scar environment.

REFERENCES: ¹ Geller HM & Fawcett JW (2002) *Exp Neurol* **172**:125-36. ² Phillips JB et al, (2004) *Biomaterials* **25**:2769-79. ³ DK Cullen, CM Simon & MC LaPlaca (2007) *Brain Res* **1158**:103-115.

ACKNOWLEDGEMENTS: This research was funded by the Wellcome Trust.